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TRANSMITTAL FORM

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TRANSMITTAL FORM (to be used for all correspondence after initial filing)	Application Number	09/884,455	
	Filing Date	June 18, 2001	
	First Named Inventor	Michael HOUGHTON	
	Art Unit	1652	
	Examiner Name	W. Moore	
Total Number of Pages in This Submission	73	Attorney Docket Number	223002010004

ENCLOSURES (Check all that apply)

<input checked="" type="checkbox"/> Fee Transmittal Form (1 page in duplicate) <input type="checkbox"/> Fee Attached <input checked="" type="checkbox"/> Amendment/Reply (17 pages) <input type="checkbox"/> After Final <input checked="" type="checkbox"/> (2) Affidavits/declaration(s) of Amy J. Weiner (28 pages) <input checked="" type="checkbox"/> Extension of Time Request (1 page) <input type="checkbox"/> Express Abandonment Request <input type="checkbox"/> Information Disclosure Statement <input type="checkbox"/> Certified Copy of Priority Document(s) <input type="checkbox"/> Reply to Missing Parts/Incomplete Application <input type="checkbox"/> Reply to Missing Parts under 37 CFR 1.52 or 1.53	<input type="checkbox"/> Drawing(s) <input type="checkbox"/> Licensing-related Papers <input type="checkbox"/> Petition <input type="checkbox"/> Petition to Convert to a Provisional Application <input type="checkbox"/> Power of Attorney, Revocation Change of Correspondence Address <input type="checkbox"/> Terminal Disclaimer <input type="checkbox"/> Request for Refund <input type="checkbox"/> CD, Number of CD(s) _____ <input type="checkbox"/> Landscape Table on CD	<input type="checkbox"/> After Allowance Communication to TC <input type="checkbox"/> Appeal Communication to Board of Appeals and Interferences <input type="checkbox"/> Appeal Communication to TC (Appeal Notice, Brief, Reply Brief) <input type="checkbox"/> Proprietary Information <input type="checkbox"/> Status Letter <input checked="" type="checkbox"/> Other Enclosure(s) (please identify below): 1. Comparison Chart (1 page) 2. Copy of Preliminary Amendment dated 2/11/02 (18 pages) 3. Second Request for Corrected Filing Receipt (2 pages) 4. Copy of Marked-up Filing Receipt (3 pages) 5. Return receipt postcard
Remarks		

SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT

Firm Name	MORRISON & FOERSTER LLP (Customer No. 25226)		
Signature			
Printed name	Shantanu Basu		
Date	October 12, 2005	Reg. No.	43,318

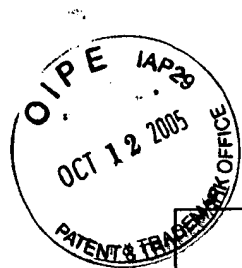
I hereby certify that this correspondence is being deposited with the U.S. Postal Service as Express Mail, Airbill No. EV544978669US, in an envelope addressed to: MS Amendment, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450, on the date shown below.

Dated: October 12, 2005

Signature: (Martina Placid)



1017 patent	'455 application
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Tamara Alcaraz

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In the application of:

Michael HOUGHTON et al.

Serial No.: 09/884,455

Filing Date: June 18, 2001

For: HEPATITIS C VIRUS PROTEASE

Examiner: To Be Assigned

Group Art Unit: 1652

PRELIMINARY AMENDMENT

Box Missing Parts
Assistant Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Preliminary to examination to examination of the above-referenced application, please
amend the specification as follows:

AMENDMENTS

In the Specification:

Please replace the entire section entitled "Brief Description of the Drawings" on page 3, lines 6 through 24:

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the sequence of HCV protease (SEQ ID NO: 69 and SEQ ID NO:70).

Figure 2 shows the polynucleotide sequence and deduced amino acid sequence of the clone C20c (SEQ ID NO: 71 and SEQ ID NO: 72).

Figure 3 shows the polynucleotide sequence and deduced amino acid sequence of the clone C26d (SEQ ID NO: 73 and SEQ ID NO: 74).

Figure 4 shows the polynucleotide sequence and deduced amino acid sequence of the clone C8h (SEQ ID NO: 75 and SEQ ID NO: 76).

Figure 5 shows the polynucleotide sequence and deduced amino acid sequence of the clone C7f (SEQ ID NO: 77 and SEQ ID NO: 78).

Figure 6 shows the polynucleotide sequence and deduced amino acid sequence of the clone C31 (SEQ ID NO: 79 and SEQ ID NO: 80).

Figure 7 shows the polynucleotide sequence and deduced amino acid sequence of the clone C35 (SEQ ID NO: 81 and SEQ ID NO: 82).

Figure 8 shows the polynucleotide sequence and deduced amino acid sequence of the clone C33c (SEQ ID NO: 83 and SEQ ID NO: 84).

Figure 9 schematically illustrates assembly of the vector C7fC2OcC300C200.

Figure 10 shows the sequence of vector cf1SODp600 (SEQ ID NO: 85 and SEQ ID NO: 86).

Please replace the paragraph starting on page 6, line 23 with the following:

The term "HCV protease" refers to an enzyme derived from HCV which exhibits proteolytic activity, specifically the polypeptide encoded in the NS3 domain of the HCV

genome. At least one strain of HCV contains a protease believed to be substantially encoded by or within the following sequence:

Arg Arg Gly Arg Glu Ile Leu Leu Gly Pro	10
Ala Asp Gly Met Val Ser Lys Gly Trp Arg	20
Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln	30
Gln Thr Arg Gly Leu Leu Gly Cys Ile Ile	40
Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln	50
Val Glu Gly Glu Val Gln Ile Val Ser Thr	60
Ala Ala Gln Thr Phe Leu Ala Thr Cys Ile	70
Asn Gly Val Cys Trp Thr Val Tyr <u>His</u> Gly	80
Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys	90
Gly Pro Val Ile Gln Met Tyr Thr Asn Val	100
Asp Gln <u>Asp</u> Leu Val Gly Trp Pro Ala Ser	110
Gln Gly Thr Arg Ser Leu Thr Pro Cys Thr	120
Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr	130
Arg His Ala Asp Val Ile Pro Val Arg Arg	140
Arg Gly Asp Ser Arg Gly Ser Leu Leu Ser	150
Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser	160
<u>Ser</u> Gly Gly Pro Leu Leu Cys Pro Ala Gly	170
His Ala Val Gly Ile Phe Arg Ala Ala Val	180
Cys Thr Arg Gly Val Ala Lys Ala Val Asp	190
Phe Ile Pro Val Glu Asn Leu Glu Thr Thr	200
Met Arg (SEQ ID NO:1)	202

Please replace the paragraph on page 8, starting on line 18 with the following:

TABLE 1: Alignment of Active Residues by Sequence

Protease	His	Asp	Ser
HCV	CWTVYHGAG(SEQ ID NO:2)	DODLGWPAP(SEQ ID NO:3)	LKGSSGGPL(SEQ ID NO:4)
Yellow Fever	FHTMWHVTR(SEQ ID NO:5)	KEDLVAYGG(SEQ ID NO:6)	PSGTSGSPI(SEQ ID NO:7)
West Nile Fever	FHTLWHTTK(SEQ ID NO:8)	KEDRLCYGG(SEQ ID NO:9)	PTGTSGSPI(SEQ ID NO:10)
Murray Valley	FHTLWHTTR(SEQ ID NO:11)	KEDRVTYGG(SEQ ID NO:12)	PIGTSGSPI(SEQ ID NO:13)
Kunjin Virus	FHTLWHTTK(SEQ ID NO:14)	KEDRLCYGG(SEQ ID NO:15)	PTGTSGSPI(SEQ ID NO:16)

Please replace the paragraph on page 9, starting on line 9 with the following:

TABLE 2: Alignment of Active Residues by Structure

Protease	His	Asp	Ser
<i>S. griseaus</i> A	TAGHC(SEQ ID NO:17)	NNDYGII(SEQ ID NO:18)	GDSGGSL(SEQ ID NO:19)
α -Lytic protease	TAGHC(SEQ ID NO:20)	GNDRAWV(SEQ ID NO:21)	GDSGGSW(SEQ ID NO:22)
Bovine Trypsin	SAAHC(SEQ ID NO:23)	NNDIMLI(SEQ ID NO:24)	GDSGGPV(SEQ ID NO:25)
Chymotrypsin	TAAHC(SEQ ID NO:26)	NNDITLL(SEQ ID NO:27)	GDSGGPL(SEQ ID NO:28)
Elastase	TAAHC(SEQ ID NO:29)	GYDIALI(SEQ ID NO:30)	GDSGGPL(SEQ ID NO:31)
HCV	TVYHG(SEQ ID NO:32)	SSDLYLV(SEQ ID NO:33)	GSSGGPL(SEQ ID NO:34)

Please replace the paragraph on page 13 starting at line 11 with the following:

A presently preferred expression system employs the ubiquitin leader as the fusion partner. Copending application USSN 7/390,599 filed 7 August 1989 disclosed vectors for high expression of yeast ubiquitin fusion proteins. Yeast ubiquitin provides a 76 amino acid polypeptide which is automatically cleaved from the fused protein upon expression. The ubiquitin amino acid sequence is as follows:

Gln Ile Phe Val Lys Thr Leu Thr Gly Lys Thr Ile Thr
 Leu Glu Val Glu Ser Ser Asp Thr Ile Asp Asn Val Lys
 Ser Lys Ile Gln Asp Lys Glu Gly Ile Pro Pro Asp Gln
 Gln Arg Leu Ile Phe Ala Gly Lys Gln Leu Glu Asp Gly
 Arg Thr Leu Ser Asp Tyr Asn Ile Gln Lys Glu Ser Thr
 Leu His Leu Val Leu Arg Leu Arg Gly Gly (SEQ ID NO:35)

Please replace the paragraph on page 21 starting at line 4 with the following:

In the absence of this protease activity, the HCV polyprotein should remain in its unprocessed form, and thus render the virus noninfectious. Thus, the protein is useful for assaying pharmaceutical agents for control of HCV, as compounds which inhibit the protease activity sufficiently will also inhibit viral infectivity. Such inhibitors may take the form of organic compounds, particularly compounds which mimic the cleavage site of HCV recognized by the protease. Three of the putative cleavage sites of the HCV polyprotein have the following amino acid sequences:

Val-Ser-Ala-Arg-Arg // Gly-Arg-Glu-Ile-Leu-Leu-Gly (SEQ ID NO:36)

Ala-Ile-Leu-Arg-Arg // His-Val-Gly-Pro- (SEQ ID NO:88)

Val-Ser-Cys-Gln-Arg // Gly-Tyr- (SEQ ID NO:89)

Please replace the paragraph on page 24 starting at line 16 with the following:

First, DNA isolated from pSODCF1 was treated with BamHI and EcoRI, and the following linker was ligated into the linear DNA created by the restriction enzymes:

GAT CCT GGA ATT CTG ATA AGA CCT TAA GAC TAT TTT AA (SEQ ID NO:37)

Please replace the paragraph on page 25 starting on line 1 with the following:

Three separate expression vectors, pcflAB, pcflCD, and pcflEF were created by ligating three new linkers, AB, CD, and EF to a BamHI-EcoRI fragment derived by digesting to completion the vector pSODF1 with EcoRI and BamHI, followed by treatment with alkaline phosphatase. The linkers were created from six oligomers, A, B, C, D, E, and F. Each oligomer was phosphorylated by treatment with kinase in the presence of ATP prior to annealing to its complementary oligomer. The sequences of the synthetic linkers were the following:

<u>Name</u>	<u>DNA Sequence (5' to 3')</u>
A	GATC CTG AAT TCC TGA TAA(SEQ ID NO:38)
B	GAC TTA AGG ACT ATT TTA A(SEQ ID NO:39)
C	GATC CGA ATT CTG TGA TAA(SEQ ID NO:40)
D	GCT TAA GAC ACT ATT TTA A(SEQ ID NO:41)
E	GATC CTG GAA TTC TGA TAA(SEQ ID NO:42)
F	GAC CTT AAG ACT ATT TTA A(SEQ ID NO:43)

Please replace the paragraph on page 27 starting on line 2 with the following:

Clone C33c was isolated using a hybridization probe having the following sequence:

5' ATC AGG ACC GGG GTG AGA ACA ATT ACC ACT 3' (SEQ ID NO:44)

Please replace the paragraph on page 27 starting on line 7 with the following:

Clone 35 was isolated by screening with a synthetic polynucleotide having the sequence:

5' AAG CCA CCG TGT GCG CTA GGG CTC AAG CCC 3' (SEQ ID NO:45)

Please replace the paragraph on page 27 starting on line 20 with the following:

Clone 7f was isolated using a probe having the sequence:

5'-AGC AGA CAA GGG GCC TCC TAG GGT GCA TAA T-3' (SEQ ID NO:46)

Please replace the paragraph on page 27 starting on line 24 with the following:

Clone C20c is isolated using a probe having the following sequence:

5'-TGC ATC AAT GGG GTG TGC TGG-3' (SEQ ID NO:47)

Please replace the paragraph on page 28 starting on line 6 with the following:

Clone 8h was isolated using a probe based on the sequence of nucleotides in clone 33c.

The nucleotide sequence of the probe was

5'-AGA GAC AAC CAT GAG GTC CCC GGT GTT C-3' (SEQ ID NO:48)

Please replace the paragraph on page 28 starting on line 11 with the following:

Clone C26d is isolated using a probe having the following sequence:

5'-CTG TTG TGC CCC GCG GCA GCC-3' (SEQ ID NO:49)

Please replace the paragraph on page 30 starting on line 8 with the following:

This vector contains a full-length HCV protease coding sequence fused to the FLAG sequence, Hopp et al. (1988) Biotechnology 6: 1204-1210. PCR was used to produce a HCV protease gene with special restriction ends for cloning ease. Plasmid p500 was digested with

EcoRI and NdeI to yield a 900 bp fragment. This fragment and two primers were used in a polymerase chain reaction to introduce a unique BglII site at amino acid 1009 and a stop codon with a SalI site at amino acid 1262 of the HCV-1, as shown in Figure 17 of WO/11089, published 4 October 1990. The sequence of the primers is as follows:

5' CCC GAG CAA GAT CTC CCG GCC C 3' (SEQ ID NO:50)

and

5' CCC GGC TGC ATA AGC AGT CGA CTT GGA 3' (SEQ ID NO:51)

After 30 cycles of PCR, the reaction was digested with BglII and SalI, and the 710 bp fragment was isolated. This fragment was annealed and ligated to the following duplex:

MetAspTyrLysAspAspAspAspLysGlyArgGlu
CATGGACTACAAAGACGATGACGATAAAGGCCGGGAG (SEQ ID NO: 52)
CTGATGTTTCTGCTACTGCTATTTCCGGCCCTCTAG (SEQ ID NO: 90)

The duplex encodes the FLAG sequence, and initiator methionine, and a 5' NcoI restriction site. The resulting NcoI/SalI fragment was ligated into a derivative of pCF1.

Please replace the paragraph on page 34 starting on line 4 with the following:

This vector contains HCV sequence, which includes the wild-type full-length HCV protease coding sequence, fused at the 5' end to a SOD coding sequence. Two fragments, a 441 bp EcoRI/BglII fragment from clone 11b and a 1471 bp BglIII/EcoRI fragment from expression vector P500, were used to reconstruct a wild-type, full-length HCV protease coding sequence. These two fragments were ligated together with an EcoRI digested pS356 vector to produce an expression cassette. The expression cassette encodes the ADH2/GAPDH hybrid yeast promoter, human SOD, the HCV protease, and a GAPDH transcription terminator. The resulting vector was digested with BamHI and a 4052 bp fragment was isolated. This fragment was ligated to the BamHI digested pAB24 vector to produce p650. p650 expresses a polyprotein containing, from its amino terminal end, amino acids 1-154 of hSOD, an oligopeptide -Asn-Leu-Gly-Ile-Arg- (SEQ ID NO:87), and amino acids 819 to 1458 of HCV-1, as shown in Figure 17 of WO 90/11089, published 4 October 1990.

Please replace the paragraph on page 34 starting on line 17 with the following:

Clone 11b was isolated from the genomic library of HCV cDNA, ATCC accession no. 40394, as described above in Example 3A, using a hybridization probe having the following sequence:

5' CAC CTA TGT TTA TAA CCA TCT CAC TCC TCT 3' (SEQ ID NO:54).

This procedure is also described in EPO Pub. No. 318 216, Example IV.A.17.

Please replace the paragraph on page 34 starting on line 22 through page 35, line 12 with the following:

The vector pS3EF, which is a pBR322 derivative, contains the ADH2/GAPDH hybrid yeast promoter upstream of the human superoxide dismutase gene, an adaptor, and a downstream yeast effective transcription terminator. A similar expression vector containing these control elements and the superoxide dismutase gene is described in Cousens et al. (1987) Gene 61: 265, and in copending application EPO 196,056, published October 1, 1986. pS3EF, however, differs from that in Cousens et al. in that the heterologous proinsulin gene and the immunoglobulin hinge are deleted, and Gln₁₅₄ of SOD is followed by an adaptor sequence which contains an EcoRI site. The sequence of the adaptor is:

5' AAT TTG GGA ATT CCA TAA TTA ATT AAG 3' (SEQ ID NO:55)

3' AC CCT TAA GGT ATT AAT TAA TTC AGCT 5' (SEQ ID NO:56)

The EcoRI site facilitates the insertion of heterologous sequences. Once inserted into pS3EF, a SOD fusion is expressed which contains an oligopeptide that links SOD to the heterologous sequences. pS3EF is exactly the same as pS356 except that pS356 contains a different adaptor. The sequence of the adaptor is shown below:

5' AAT TTG GGA ATT CCA TAA TGA G 3' (SEQ ID NO:57)

3' AC CCT TAA GGT ATT ACT CAG CT 5' (SEQ ID NO:58)

pS356, ATCC accession no. 67683, is deposited as set forth below.

Please replace the paragraph on page 37 starting at line 4 with the following:

Four synthetic DNA fragments were annealed and ligated** together to create a EcoRI/SacI Yellow Fever leader, which was ligated to a EcoRI/SacI digested pGEM®-3Z vector from Promega®. The sequence of the four fragments are listed below:

YFK-1:

5' AAT TCG TAA ATC CTG TGT GCT AAT TGA GGT GCA TTG GTC TGC
AAA TCG AGT TGC TAG GCA ATA AAC ACA TT 3' (SEQ ID NO:59)

YFK-2:

5' TAT TGC CTA GCA ACT CGA TTT GCA GAC CAA TGC ACC TCA ATT AGC
ACA CAG GAT TTA CG 3' (SEQ ID NO:60)

YFK-3:

5' TGG ATT AAT TIT AAT CGT TCG TTG AGC GAT TAG CAG AGA ACY GAC
CAG AAC ATG TCT GAG CT 3' (SEQ ID NO:61)

YFK-4:

5' CAG ACA TGT TCT GGT CAG TTC TCT GCT AAT CGC TCA ACG AAC
GAT TAA AAT TAA TCC AAA TGT GTT 3' (SEQ ID NO:62)

REMARKS

Applicant submits herewith Sequence Listings of nucleotide sequences disclosed in this application in accordance with 37 C.F.R. §§ 1.821-1.825. The Specification is amended on pages 3, 6, 8, 9, 13, 21, 24, 25, 27, 28, 30, 34 and 37, to conform with the requirements of Sequence disclosures.

Attached hereto is a marked-up version of the changes made to the specification by the current amendment. The attached page is captioned "Version with markings to show changes made".

In the unlikely event that the transmittal letter is separated from this document and the Patent Office determines that an extension and/or other relief is required, applicant petitions for any required relief including extensions of time and authorizes the Assistant Commissioner to charge the cost of such petitions and/or other fees due in connection with the filing of this document to Deposit Account No. 03-1952 referencing docket no. 223002010004.

Respectfully submitted,

Dated: February 11, 2002

By:



Cara M. Coburn
Registration No. 46,631

for

Gladys H. Monroy
Registration No. 32,430

Morrison & Foerster LLP
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Palo Alto, California 94304-1018
Telephone: (650) 813-4218
Facsimile: (650) 494-0792

VERSION WITH MARKINGS TO SHOW CHANGES MADE

In the Specification:

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BRIEF DESCRIPTION OF THE DRAWINGS

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Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln	50
Val Glu Gly Glu Val Gln Ile Val Ser Thr	60
Ala Ala Gln Thr Phe Leu Ala Thr Cys Ile	70
Asn Gly Val Cys Trp Thr Val Tyr His Gly	80
Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys	90
Gly Pro Val Ile Gln Met Tyr Thr Asn Val	100
Asp Gln Asp Leu Val Gly Trp Pro Ala Ser	110
Gln Gly Thr Arg Ser Leu Thr Pro Cys Thr	120
Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr	130
Arg His Ala Asp Val Ile Pro Val Arg Arg	140
Arg Gly Asp Ser Arg Gly Ser Leu Leu Ser	150
Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser	160
Ser Gly Gly Pro Leu Leu Cys Pro Ala Gly	170
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Murray Valley	FHTLWHTTR(SEQ ID NO:11)	KEDRVTYGG(SEQ ID NO:12)	PIGTSGSPI(SEQ ID NO:13)
Kunjin Virus	FHTLWHTTK(SEQ ID NO:14)	KEDRLCYGG(SEQ ID NO:15)	PTGTSGSPI(SEQ ID NO:16)

Please replace the paragraph on page 9, starting on line 9 with the following:

TABLE 2: Alignment of Active Residues by Structure

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Elastase	<u>TAAHC(SEQ ID NO:29)</u>	<u>GYDIALI(SEQ ID NO:30)</u>	<u>GDSGGPL(SEQ ID NO:31)</u>
HCV	<u>TVYHG(SEQ ID NO:32)</u>	<u>SSDLYLV(SEQ ID NO:33)</u>	<u>GSSGGPL(SEQ ID NO:34)</u>

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A presently preferred expression system employs the ubiquitin leader as the fusion partner. Copending application USSN 7/390,599 filed 7 August 1989 disclosed vectors for high expression of yeast ubiquitin fusion proteins. Yeast ubiquitin provides a 76 amino acid polypeptide which is automatically cleaved from the fused protein upon expression. The ubiquitin amino acid sequence is as follows:

Gln Ile Phe Val Lys Thr Leu Thr Gly Lys Thr Ile Thr
 Leu Glu Val Glu Ser Ser Asp Thr Ile Asp Asn Val Lys
 Ser Lys Ile Gln Asp Lys Glu Gly Ile Pro Pro Asp Gln
 Gln Arg Leu Ile Phe Ala Gly Lys Gln Leu Glu Asp Gly
 Arg Thr Leu Ser Asp Tyr Asn Ile Gln Lys Glu Ser Thr
 Leu His Leu Val Leu Arg Leu Arg Gly Gly (SEQ ID NO:35)

Please replace the paragraph on page 21 starting at line 4 with the following:

In the absence of this protease activity, the HCV polyprotein should remain in its unprocessed form, and thus render the virus noninfectious. Thus, the protease is useful for assaying pharmaceutical agents for control of HCV, as compounds which inhibit the protease activity sufficiently will also inhibit viral infectivity. Such inhibitors may take the form of organic compounds, particularly compounds which mimic the cleavage site of HCV recognized by the protease. Three of the putative cleavage sites of the HCV polyprotein have the following amino acid sequences:

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Val-Ser-Cys-Gln-Arg // Gly-Tyr- (SEQ ID NO:89)

Please replace the paragraph on page 24 starting at line 16 with the following:

First, DNA isolated from pSODCF1 was treated with BamHI and EcoRI, and the following linker was ligated into the linear DNA created by the restriction enzymes:

GAT CCT GGA ATT CTG ATA AGA CCT TAA GAC TAT TTT AA (SEQ ID NO:37)

Please replace the paragraph on page 25 starting on line 1 with the following:

Three separate expression vectors, pcf1AB, pcf1CD, and pcf1EF were created by ligating three new linkers, AB, CD, and EF to a BamHI-EcoRI fragment derived by digesting to completion the vector pSODF1 with EcoRI and BamHI, followed by treatment with alkaline phosphatase. The linkers were created from six oligomers, A, B, C, D, E, and F. Each oligomer was phosphorylated by treatment with kinase in the presence of ATP prior to annealing to its complementary oligomer. The sequences of the synthetic linkers were the following:

Name	DNA Sequence (5' to 3')
A	GATC CTG AAT TCC TGA TAA(SEQ ID NO:38)
B	GAC TTA AGG ACT ATT TTA A(SEQ ID NO:39)
C	GATC CGA ATT CTG TGA TAA(SEQ ID NO:40)
D	GCT TAA GAC ACT ATT TTA A(SEQ ID NO:41)
E	GATC CTG GAA TTC TGA TAA(SEQ ID NO:42)
F	GAC CTT AAG ACT ATT TTA A(SEQ ID NO:43)

Please replace the paragraph on page 27 starting on line 2 with the following:

Clone C33c was isolated using a hybridization probe having the following sequence:

5' ATC AGG ACC GGG GTG AGA ACA ATT ACC ACT 3' (SEQ ID NO:44)

Please replace the paragraph on page 27 starting on line 7 with the following:

Clone 35 was isolated by screening with a synthetic polynucleotide having the sequence:

5' AAG CCA CCG TGT GCG CTA GGG CTC AAG CCC 3' (SEQ ID NO:45)

Please replace the paragraph on page 27 starting on line 20 with the following:

Clone 7f was isolated using a probe having the sequence:

5'-AGC AGA CAA GGG GCC TCC TAG GGT GCA TAA T-3' (SEQ ID NO:46)

Please replace the paragraph on page 27 starting on line 24 with the following:

Clone C20c is isolated using a probe having the following sequence:

5'-TGC ATC AAT GGG GTG TGC TGG-3' (SEQ ID NO:47)

Please replace the paragraph on page 28 starting on line 6 with the following:

Clone 8h was isolated using a probe based on the sequence of nucleotides in clone 33c.

The nucleotide sequence of the probe was

5'-AGA GAC AAC CAT GAG GTC CCC GGT GTT C-3' (SEQ ID NO:48)

Please replace the paragraph on page 28 starting on line 11 with the following:

Clone C26d is isolated using a probe having the following sequence:

5'-CTG TTG TGC CCC GCG GCA GCC-3' (SEQ ID NO:49)

Please replace the paragraph on page 30 starting on line 8 with the following:

This vector contains a full-length HCV protease coding sequence fused to the FLAG sequence, Hopp et al. (1988) Biotechnology 6: 1204-1210. PCR was used to produce a HCV protease gene with special restriction ends for cloning ease. Plasmid p500 was digested with EcoRI and NdeI to yield a 900 bp fragment. This fragment and two primers were used in a polymerase chain reaction to introduce a unique BglII site at amino acid 1009 and a stop codon

with a Sall site at amino acid 1262 of the HCV-1, as shown in Figure 17 of WO/11089, published 4 October 1990. The sequence of the primers is as follows:

5' CCC GAG CAA GAT CTC CCG GCC C 3' (SEQ ID NO:50)

and

5' CCC GGC TGC ATA AGC AGT CGA CTT GGA 3' (SEQ ID NO:51)

After 30 cycles of PCR, the reaction was digested with BglII and Sall, and the 710 bp fragment was isolated. This fragment was annealed and ligated to the following duplex:

MetAspTyrLysAspAspAspLysGlyArgGlu
CATGGACTACAAAGACGATGACGATAAAGGCCGGGAG (SEQ ID NO:52)
CTGATGTTTCTGCTACTGCTATTTCCGGCCCTCTAG (SEQ ID NO:90)

The duplex encodes the FLAG sequence, and initiator methionine, and a 5' NcoI restriction site. The resulting NcoI/Sall fragment was ligated into a derivative of pCF1.

Please replace the paragraph on page 34 starting on line 4 with the following:

This vector contains HCV sequence, which includes the wild-type full-length HCV protease coding sequence, fused at the 5' end to a SOD coding sequence. Two fragments, a 441 bp EcoRI/BglII fragment from clone 11b and a 1471 bp BglIII/EcoRI fragment from expression vector P500, were used to reconstruct a wild-type, full-length HCV protease coding sequence. These two fragments were ligated together with an EcoRI digested pS356 vector to produce an expression cassette. The expression cassette encodes the ADH2/GAPDH hybrid yeast promoter, human SOD, the HCV protease, and a GAPDH transcription terminator. The resulting vector was digested with BamHI and a 4052 bp fragment was isolated. This fragment was ligated to the BamHI digested pAB24 vector to produce p650. p650 expresses a polyprotein containing, from its amino terminal end, amino acids 1-154 of hSOD, an oligopeptide -Asn-Leu-Gly-Ile-Arg- (SEQ ID NO:87), and amino acids 819 to 1458 of HCV-1, as shown in Figure 17 of WO 90/11089, published 4 October 1990.

Please replace the paragraph on page 34 starting on line 17 with the following:

Clone 11b was isolated from the genomic library of HCV cDNA, ATCC accession no. 40394, as described above in Example 3A, using a hybridization probe having the following sequence:

5' CAC CTA TGT TTA TAA CCA TCT CAC TCC TCT 3' (SEQ ID NO:54).

This procedure is also described in EPO Pub. No. 318 216, Example IV.A.17.

Please replace the paragraph on page 34 starting on line 22 through page 35, line 12 with the following:

The vector pS3EF, which is a pBR322 derivative, contains the ADH2/GAPDH hybrid yeast promoter upstream of the human superoxide dimutase gene, an adaptor, and a downstream yeast effective transcription terminator. A similar expression vector containing these control elements and the superoxide dismutase gene is described in Cousens et al. (1987) Gene 61: 265, and in copending application EPO 196,056, published October 1, 1986. pS3EF, however, differs from that in Cousens et al. in that the heterologous proinsulin gene and the immunoglobulin hinge are deleted, and Gln₁₅₄ of SOD is followed by an adaptor sequence which contains an EcoRI site. The sequence of the adaptor is:

5' AAT TTG GGA ATT CCA TAA TTA ATT AAG 3' (SEQ ID NO:55)

3' AC CCT TAA GGT ATT AAT TAA TTC AGCT 5' (SEQ ID NO:56)

The EcoRI site facilitates the insertion of heterologous sequences. Once inserted into pS3EF, a SOD fusion is expressed which contains an oligopeptide that links SOD to the heterologous sequences. pS3EF is exactly the same as pS356 except that pS356 contains a different adaptor. The sequence of the adaptor is shown below:

5' AAT TTG GGA ATT CCA TAA TGA G 3' (SEQ ID NO:57)

3' AC CCT TAA GGT ATT ACT CAG CT 5' (SEQ ID NO:58)

pS356, ATCC accession no. 67683, is deposited as set forth below.

Please replace the paragraph on page 37 starting at line 4 with the following:

Four synthetic DNA fragments were annealed and ligated** together to create a EcoRI/SacI Yellow Fever leader, which was ligated to a EcoRI/SacI digested pGEM®-3Z vector from Promega®. The sequence of the four fragments are listed below:

YFK-1:

5' AAT TCG TAA ATC CTG TGT GCT AAT TGA GGT GCA TTG GTC TGC
AAA TCG AGT TGC TAG GCA ATA AAC ACA TT 3' (SEQ ID NO:59)

YFK-2:

5' TAT TGC CTA GCA ACT CGA TTT GCA GAC CAA TGC ACC TCA ATT AGC
ACA CAG GAT TTA CG 3' (SEQ ID NO:60)

YFK-3:

5' TGG ATT AAT TIT AAT CGT TCG TTG AGC GAT TAG CAG AGA ACY GAC
CAG AAC ATG TCT GAG CT 3' (SEQ ID NO:61)

YFK-4:

5' CAG ACA TGT TCT GGT CAG TTC TCT GCT AAT CGC TCA ACG AAC
GAT TAA AAT TAA TCC AAA TGT GTT 3' (SEQ ID NO:62)